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Product Number: AQ-11949LF

AQuora® 650-NHS Ester

AQuora® 650-NHS Ester is an amine-reactive fluorophore engineered with SuperHydrophilic™ technology designed to improve solubility during labeling and of the dye labeled conjugate. As a result, the dye labeled conjugates made with AQuora® 650-NHS Ester yield enhanced signal and signal-to-noise ratios in fluorescence-based applications including fluorescent western blotting, fluorescence-based microscopy, flow cytometry, and cell-based assays. AQuora® 650-NHS Ester is a cyanine-based dye with photophysical properties comparable to Alexa Fluor® 647, Cy5, DyLight® 650, DyLight® 650 4x PEG.

This AQuora® 650-NHS Ester dye is activated with a N-hydroxysuccinimide (NHS) ester group which reacts with free primary amines, such as the amines on the side chain of lysines, forming a stable amide bond.

Alexa Fluor® and DyLight® are registered trademarks of Thermo Fisher Scientific

Product Number:	**AQ-11949LF**
Unit Size:	1 mg
Format:	Solid
Concentration:	Refer to the lot-specific Certificate of Analysis for the dye concentration.
Storage and Handling:	Store at -20°C. Protect from light. May ship at ambient temperatures.
Instructions for Use:	Equilibrate dye solution to room temperature before opening. For its use in labeling reactions, follow the guidelines in the example protocol below.

Label Information	**AQuora® 650**
Reactive Group:	NHS Ester
Molecular Weight:	1710 g/mol
Extinction Coefficient (-M-cm):	250,000 $cm^{-1}M^{-1}$
A_{280} Correction Factor:	0.037
Excitation / Emission Maximum (nm):	654 nm/674 nm ± 4nm
Spectrally Similar Dyes:	Alexa Fluor® 647, Cy5, DyLight® 650, DyLight® 650 4x PEG

Example Conjugation Reaction

AQuora® 650-NHS Ester is an amine-reactive dye that can be conjugated to many proteins or peptides. The NHS ester reacts with primary amines to form a stable, covalent amide bond. NHS ester reagents are moisture sensitive, and the dyes are light and oxygen sensitive; therefore, store the dye solution protected from light and blanketed with inert gas, such as nitrogen or argon.

General Protocol for Dye Labeling of Antibody

Prepare the antibody for labeling

The optimal antibody concentration for labeling is 5-10 mg/ml. When using lower antibody concentrations, the amount of dye added to the labeling reaction may have to be increased to obtain an optimal degree of labeling (DOL) or fluor-to-protein (F/P) labeling ratio.

1. If any precipitate (cloudiness or aggregate material) is visibly present in the antibody solution, filter it through a 0.2 µm syringe filter before use.

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2. Dialyze the filtered antibody solution for 2 hours against 50 mM sodium borate, 50 mM sucrose, pH 8.5 in a volume 200 times the sample volume using a Slide-A-Lyzer® G2 50K MWCO dialysis cassette or the equivalent. Repeat the dialysis one more time with fresh dialysis buffer for an additional 2 hours. For small volumes, an Amicon Ultracel 50K MWCO centrifugal filter can be used to buffer exchange and concentrate the antibody stock solution. Several spins (4-5) of concentrating and diluting with fresh buffer may be needed to fully buffer exchange the antibody using the centrifugal filter method.

3. Determine the Antibody Concentration using the following formula. A small aliquot of the antibody solution may have to be diluted with buffer to measure the A280 nm in order to keep the absorbance within the linear range of the spectrophotometer.

   Antibody concentration (mg/ml) = $[(A_{280})$(dilution factor)] / (extinction coefficient of the antibody)

   Where the dilution factor is the amount that a small sample of the antibody solution had to be diluted to measure its absorbance. For instance, if 10 µl of the antibody solution was diluted with 90 µl of buffer to measure the A_{280} nm, then the dilution factor is 10 (because 10 µl was diluted up to 100 µl).

   And where the extinction coefficient (ε_{IgG} in $cm^{-1}M^{-1}$) is the absorbance at 280 nm of a 1 mg/ml concentration of the antibody (using a 1 cm path length). For many polyclonal antibodies, the extinction coefficient is approximately 1.4. If your antibody gives a different extinction coefficient, then replace this value with your determined value.

4. Determine amount of antibody to label (mg) using the follow formula:

   mg of antibody = (volume of antibody solution, in ml)(antibody concentration, in mg/ml)

5. Adjust the antibody concentration to approximately 10 mg/ml by adding reaction buffer or concentrating the solution as needed.

Prepare the dye stock solution

1. Bring the vial containing the dye (either supplied as a solid or as a DMAC solution) to room temperature to prevent moisture condensation upon opening. Protect from light. If the dye is supplied as a DMAC solution, use the concentration provided on the lot-specific certificate of analysis and skip to the next section.

2. If the dye is supplied as a solid, add anhydrous solvent [dimethyl sulfoxide (DMSO), dimethylacetamide (DMAC) or dimethylformamide (DMF)] and mix by vortexing. For a 10 mg/ml stock concentration, add 100 µl of anhydrous solvent to the vial containing 1 mg of reactive dye.

   To make the solvent anhydrous for this purpose, it should be treated with a molecular sieve which prior to use had been incubated at 250°C for 16 hours under vacuum to remove residual water and to fully activate the sieves, then cooled to room temperature. Reactive dyes prepared in an anhydrous solvent, protected from light, and sealed for storage at -20°C, are stable for at least 2 months. Do not store reactive dyes prepared in anhydrous solvent for long periods as stability cannot be assured. Reactive dyes dissolved in solvents that contain residual water will lose NHS ester activity due to hydrolysis.

3. Determine the molar concentration of the dye stock solution according to the following equation:

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Molar concentration of dye (M) = [(mg dye) / (ml solvent)] / (MW of dye in g/mol)

Determine the volume of dye to be used for labeling

The mole excess of dye over the antibody used in the reaction for the optimal final degree of labeling (DOL) should be analytically determined. In general, a 5-10-fold mole excess is appropriate for labeling antibodies prepared at a concentration of 5-10 mg/ml. More dilute solutions of antibody may require greater amounts of dye to obtain the same DOL.

1. Calculate the appropriate volume of dye solution to add per mg of antibody according to the following equations:

   Mole IgG = (g IgG) / (150,000 g/mol)

   where the MW of an intact polyclonal IgG antibody is approximately 150,000 g/mol. When labeling monoclonal or recombinant antibodies, antibody fragments, or other proteins, adjust the MW to match that of the protein being labeled.

   Mole excess of dye desired in reaction: _____

   Moles of label required = (mol IgG)(mol excess of dye desired)

   Volume of label needed (µl) = (mol label required) / [(M of dye solution)(10^6 µl/L)]

Reaction conditions

1. Add the calculated volume of the AQ® dye to the antibody mixture *while* gently vortexing for rapid dissolution. Cap the tube and gently vortex again for ~30 seconds.

2. Allow the conjugation reaction to gently rock for at least 1 hour at room temperature. Longer reaction times may increase the DOL and yield of the reaction; however, the time of the reaction should be controlled to obtain similar DOL's from batch to batch. A 25°C constant temperature water bath may be used with periodic mixing to enhance the reproducibility of labeling results.

Purification

The purification method should be adjusted based upon the quantity of antibody to be labeled and the reaction volume.

1. Prepare a Sephadex G-50 (or equivalent) size exclusion support in a purification buffer appropriate for the final application or use 10 mM sodium phosphate, 150 mM sodium chloride, pH 7.2.

2. Pack the hydrated G-50 support into the appropriate column ensuring that the packed gel has a volume 20 times larger than the reaction sample volume to be purified. Equilibrate the packed gel with 3 bed volumes of purification buffer.

3. Gently load the sample onto the top of the gel bed and start the UV monitoring system, if available.

4. Elute the dye-labeled protein using the purification buffer by connecting the column to a solvent reservoir of the buffer to maintain continuous flow through the column. Collect appropriately sized fractions as the flow of buffer comes through the column.

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5. Collect and pool together the early-eluting, colored fractions corresponding to the first peak which is the dye-labeled protein. The broad, second peak is the excess dye, which may be discarded as aqueous waste.

   Alternative Method: For small volumes of labeling reactions, remove excess dye from labeled antibodies using an Amicon Ultracel 50K MWCO 0.5 ml spin concentrator and wash with ~6 washes of purification buffer by repeatedly diluting and concentrating the labeled antibody.

Analysis of Labeled Antibody

The non-reacted excess dye must be completely removed for accurate quantitation of labeled antibody concentration and DOL (or the fluor-to-protein (F/P) ratio).

1. Dilute a small amount of the labeled protein in the purification buffer to measure its absorbance properties. It is important to dilute the sample enough to get the measurement of the A_{max} of the dye within the linear range of the spectrophotometer.

   Recommended dilution factor: 1:49 (which is a 1→50 dilution) to 1:99 (or 1→100)

2. Measure the absorbance at 280 nm and at the A_{max} of the dye using a 1 cm quartz cuvette.

3. Calculate the concentration of antibody using the following formulas and the values determined above:

   Concentration of diluted antibody (M) = $[A_{280}-(A_{max})(CF)] / \varepsilon_{IgG}$ at 280 nm

   Where CF is the A_{280} correction factor of the dye ($CF = A_{280nm}/A_{max}$) (see above under Label Information for the specific dye CF value), and

   ε_{IgG} at 280 nm = 205,800 $cm^{-1}M^{-1}$

   This represents the extinction coefficient for a typical polyclonal antibody. If the sample is not a polyclonal antibody, then replace this value with the extinction coefficient appropriate for the antibody or protein being labeled.

   Diluted antibody concentration (mg/ml) = (Molarity, mmol/ml)(MW_{IgG}, mg/mmol)

   Where MW_{IgG} = 150,000 mg/mmol

   Original antibody sample concentration (mg/ml) = (diluted antibody conc., in mg/ml)(DF)

   Where DF is the dilution factor used to measure the absorbance

4. Calculate the degree of labeling (in moles of dye per moles of antibody) using the following formula:

   Degree of Labeling (DOL) = $[(A_{max})(DF)] / [(\varepsilon_{dye}\ M^{-1}cm^{-1})$(antibody conc., M)]

Target Degree of Labeling

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The optimal DOL should be determined based on the antibody or protein being labeled and its intended application. For many antibodies used in immunoassays, a final DOL of 4-7 dyes per antibody, yields a maximal signal and signal-to-noise ratio.

Troubleshooting

Problem	Cause	Solution
Protein was not labeled; insufficiently labeled	The NHS Ester has hydrolyzed and is non-reactive	Prepare labeling reagent immediately before use in an anhydrous solvent. Do not prepare in an aqueous solution.
	Conjugation buffer contained primary amines that interfere with the reaction	Ensure reaction buffer is free of amines (i.e., no tris or glycine)
	Too low of molar excess in the reaction and/or protein concentration too dilute	Increase the molar fold excess of reactive dye and prepare the protein at a concentration of ~5-10 mg/ml. The more dilute the protein, the higher the molar excess of dye necessary to reach the target degree of labeling.
The degree of labeling is higher than anticipated	Insufficient removal of excess dye	Separate labeled proteins by size exclusion chromatography. Passive dialysis is not recommended for the removal of excess AQuora® reactive dyes.
Detection of AQuora® dye-labeled protein was low or absent	Instrument settings are not optimal or set for specific fluor	Ensure instrument specifications for excitation and emission are appropriate for the AQuora® fluor.
	Protein was insufficiently labeled	See solutions above
	Protein over-labeled causing fluorophore quenching and/or inhibition of binding capacity of the protein	Optimize reaction conditions for a degree of labeling appropriate for the assay